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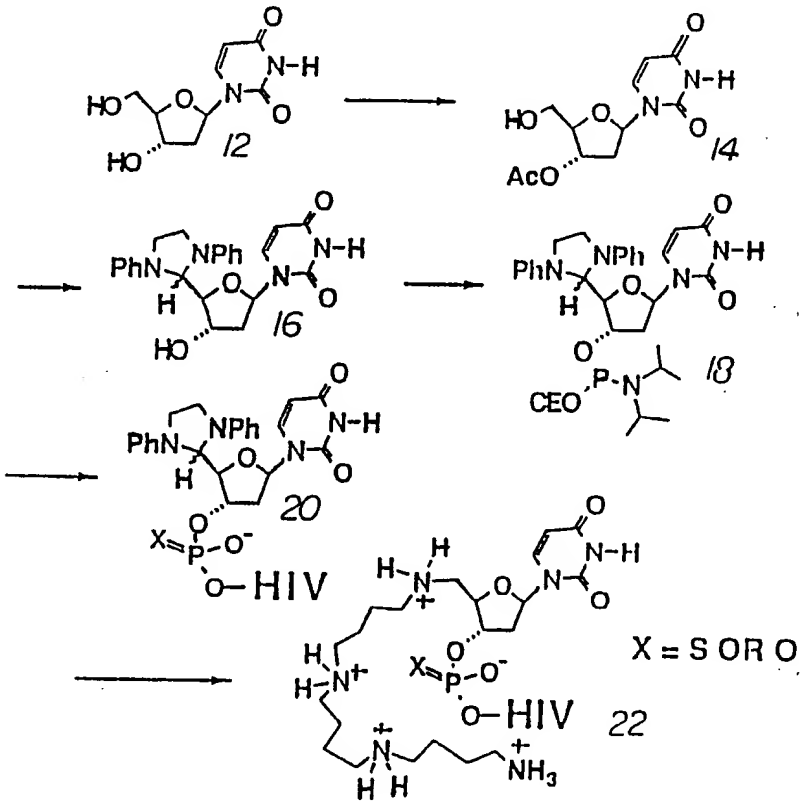
(57) Abstract

Oligonucleotide analogs are provided having improved cellular uptake and improved nuclease resistance. Modification of oligonucleotides through the attachment of nitrogenous moieties, especially polyamines, hydrazines and the like to nucleosidic portions of the analogs is disclosed. Oligonucleotides targeted at the *tat* region of HIV comprise certain preferred embodiments.



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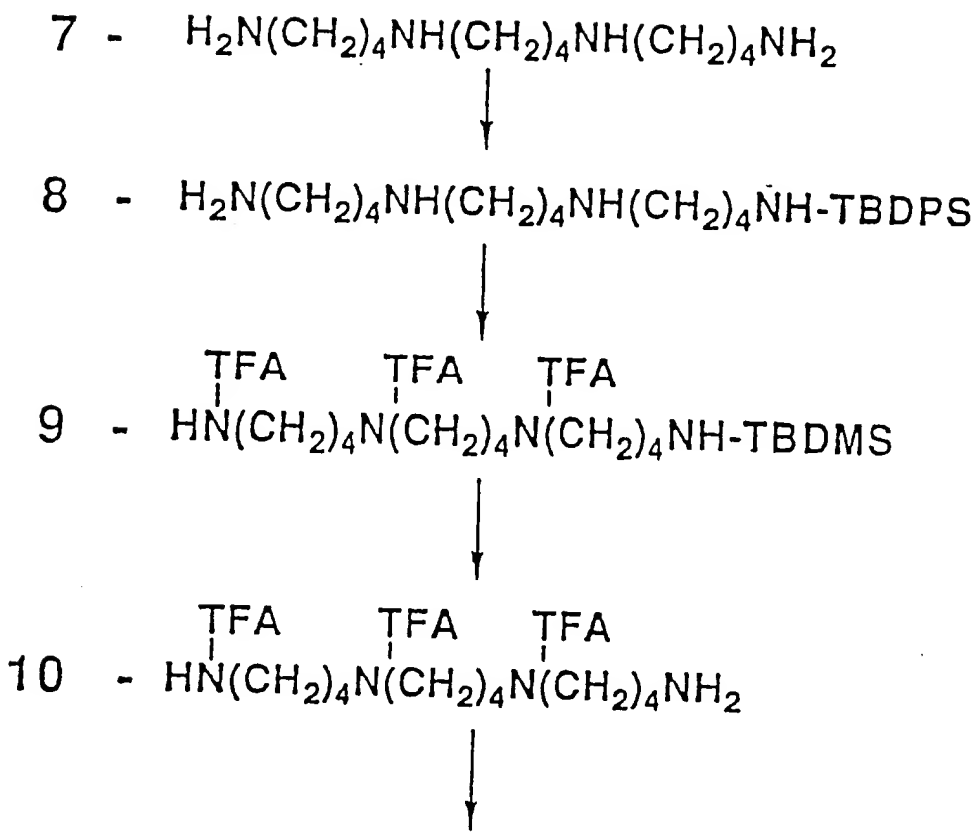
7 - $H_2N(CH_2)_4NH(CH_2)_4NH(CH_2)_4NH_2$
8 - $H_2N(CH_2)_4NH(CH_2)_4NH(CH_2)_4NH$-TBDPS
TFA TFA TFA
9 - $HN(CH_2)_4N(CH_2)_4N(CH_2)_4NH$-TBDMS
TFA TFA TFA
10 - $HN(CH_2)_4N(CH_2)_4N(CH_2)_4NH_2$
OLIGONUCLEOTIDE 6

FIGURE 1

NOVEL POLYAMINE CONJUGATED OLIGONUCLEOTIDES

FIELD OF THE INVENTION

This invention relates to the field of therapeutics, and in particular to the treatment of infection by antisense therapy. This invention also relates to the field of gene expression. Novel polyamine conjugated phosphorothioate oligonucleotides which are useful in antisense therapy are provided. These oligonucleotides have enhanced cellular uptake and consequently enhanced biological and therapeutic activity. The invention also provides methods of synthesis of novel polyamine conjugated phosphorothioate oligonucleotides.

BACKGROUND OF THE INVENTION

It is well known that most of the bodily states in mammals including most disease states, are effected by proteins. Such proteins, either acting directly or through their enzymatic functions, contribute in major proportion to many diseases in animals and man. Classical therapeutics has generally focused upon interactions with such proteins in efforts to moderate their disease causing or disease potentiating functions. Recently, however, attempts have been made to moderate the actual production of such proteins by interactions with molecules that direct their synthesis, intracellular RNA. These interactions involved the binding of complementary "antisense" oligonucleotides or their analogs to the transcellular RNA in a sequence specific fashion such as by Watson-Crick base pairing interactions.

The pharmacological activity of antisense oligonucleotides, as well as other therapeutics, depends on a number of factors that influence the effective concentration of these agents at specific intracellular targets. One important factor is the stability of the oligonucleotide in the presence of nucleases. Another key factor is the ability of antisense compounds to traverse the plasma membrane of specific cells involved in the disease process.

Cellular membranes consist of lipid protein bilayers that are freely permeable to small, nonionic, lipophilic compounds and inherently impermeable to most natural metabolites and therapeutic agents, Wilson, D.B. *Ann. Rev. Biochem.* 47:933-965 (1978). The biological and antiviral effects of natural and modified oligonucleotides in cultured mammalian cells have been well documented, so it appears that these agents can penetrate membranes to reach their intracellular targets. Uptake of antisense compounds into a variety of mammalian cells, including HL-60, Syrian Hamster fibroblast, U937, L929, CV-1, and ATH8 cells has been studied using natural oligonucleotides and nuclease resistant analogs, such as alkyl triesters, Miller, P.S., Braiterman, L.T. and Ts'O, P.O.P., *Biochemistry* 16:1988-1996 (1977); methylphosphonates, Marcus-Sekura, C.H., Woerner, A.M., Shinozuka, K. Zon, G., and Quinman, G.V., *Nuc. Acids Res.* 15:5749-5763 (1987) and Miller, P.S., McParland, K.B., Hayerman, K. and Ts'O, P.O.P., *Biochemistry* 20:1874-1880 (1981); and phosphorothioates, Ceruzzi, M. and Draper, K. *Nucleosides & Nucleotides* 8:815-818 (1989); Miller, P.S., Braiterman, L.T. and Ts'O, P.O.P. *Biochemistry* 16:1988-1996 (1977) and Loke, S.L., Stein, C., Zhang, X.H. Avigan, M., Cohen, J. and Neckers, L.M. *Curr. Top. Microbiol. Immunol.* 141:282-289 (1988).

Phosphorothioates are oligonucleotide analogs in which the oxygen atom in each phosphate linkage is replaced by a sulfur. Although the overall charge is conserved, and

they are therefore comparable in that respect to phosphodiester oligonucleotides, several properties of this class of analogs makes them more attractive than other modified compounds. These include ease of chemical synthesis, good aqueous solubility, relatively high resistance to nucleases, and the ability to form stable duplexes with complementary DNA or RNA strands. However, phosphorothioates were studied concurrently with natural compounds by Loke et al, *Proc. Natl. Acad. Sci. U.S.A.* 86:3473-3478 (1989), and while they may be useful due to their nuclease resistance, they are less efficiently internalized than their natural oligonucleotide counterparts.

Advances in nucleotide chemistry have allowed attachment of functional groups to the 3' and 5' end of the oligonucleotides to enhance cellular uptake in specific cell types. Previous studies have shown that plasmid DNA complexed with an asialoglycoprotein-poly(L-lysine) conjugate, could be targeted to hepatocytes, which contain unique cell surface receptors for galactose-terminal asialoglycoproteins, Wu, G.Y. and Wu, C.H. *Biochemistry* 27:887-892 (1988). Other groups have synthesized oligodeoxyribonucleotides that have a 5'-attached alkylating agent and a 3' attached cholesterol moiety and determined that these modified oligonucleotides were taken up into cells more efficiently than control compounds without the steroid moiety, Zon, G. in *Oligodeoxynucleotides: Antisense Inhibitors of Gene Expression* 234-247, ed. J.S. Cohen (CRC Press, Boca Raton FL, 1989). Letsinger, et al. *Proc. Natl. Acad. Sci. U.S.A.* 86:653-656 (1989), have also synthesized cholesteryl-conjugated phosphorothioates whose anti-HIV activity is significantly greater than natural oligonucleotides with the same sequence. Additional modifications include conjugation of oligonucleotides to poly(L-lysine) alone, Stevenson, M. and Iversen, P.L. *J. Gen. Virol* 70:2673-2682 (1989) and Lemaitre, M., Baynard, B. and LeBleu, B. *Proc.*

Natl. Acad. Sci. U.S.A. 84:648-652 (1987). This modification enhanced the antiviral activity of the compound studied presumably due to increased cellular uptake imparted by the polycationic poly(L-lysine).

The activity of antisense oligonucleotides previously available has not been sufficient for practical therapeutic, research or diagnostic use. The basis of this insufficiency is likely several fold i.e., (1) incomplete understanding of the secondary and tertiary structure of the targeted RNA, (2) low percentages of delivery and uptake, (3) inactivation of reactive centers by other cellular components, and (4) requirements for stoichiometric conditions for inhibition of protein production.

Enhancement of cellular uptake of antisense oligonucleotides by chemical modification would have clear advantages. Novel modifications may also lead to increased lipophilicity, greater retention, and increased distribution of the novel compounds. Increasing the concentration of oligonucleotides at specific intracellular target sites may ultimately increase the safety and efficacy of these compounds since less of the drug will be required to produce the desired effects.

Accordingly, there has been and continues to be a long-felt need for oligonucleotides and oligonucleotide analogs which are capable of effective therapeutic and diagnostic antisense use and specifically an oligonucleotide or oligonucleotide analog which is comprised of a functional group which facilitates transport into the cell and at the same time is less susceptible to nuclease activity than wild types. This long felt need has not been satisfied by prior work in the field of antisense oligonucleotide therapy and diagnostics.

SUMMARY OF THE INVENTION

In accordance with this invention, novel types of antisense oligonucleotides are provided which are modified to enhance cellular uptake. Oligonucleotides having at

least one nucleoside modified by direct attachment to a polyamine have been found to be effective in accomplishing this goal. It is preferred that the nucleoside be attached at its 5' site of the sugar moiety to the polyamine. It is preferred that the oligonucleotides in accordance with this invention be modified so as to be phosphorothioates or other backbone modified species. The oligonucleotides may preferably range in length from about 5 to about 50 nucleic acid bases. In accordance with preferred embodiments of this invention, the oligonucleotides code for the *tat* region of a HIV genome. In accordance with other preferred embodiments, the selected sequence coding for the *tat* region of the HIV genome has a thymidine at its 5' terminal end. Other preferred antisense oligonucleotide sequences include complementary sequences for herpes, papilloma and other viruses.

The modified nucleoside preferably found at the 5' end of the phosphorothioate oligonucleotide may be any pyrimidine or purine. However, preferred embodiments of this invention incorporate a modified thymidine at the 5' end.

The modified nucleosides are preferably nuclease resistant linkages of a functional group on a polyamine joined to the nucleoside at the 5' site of its sugar moiety. The polyamine functional group may comprise primary amines, hydrazines, semicarbazines, thiosemicarbazines or similar nitrogenous species. A preferred configuration of the polyamine incorporates a symmetrical carbon spacing group between each amine function with the functional group on a terminal end of the polyamine.

The linkage between the modified nucleoside and the polyamine functional group is preferably, generally unlike functional group additions presently known in the art. Rather than the usual phosphodiester linkage, the addition occurs directly at the 5' position. Not only is this linkage an improvement over the present state of the

art because it is nuclease resistant, but in addition the direct attachment of this functional group to the oligonucleotide or oligonucleotide analog confers superior cellular uptake relative to the naturally occurring oligonucleotide. This superior cellular uptake may likely be due to the neutralizing effect that the polyamine has on the negative charges of the oligonucleotide since the polyamine may be directed back along the sugar-phosphate backbone of the oligonucleotide conjugate as well as the backbone of the polyamine-oligonucleotide heteroduplex.

This invention is also directed to methods for synthesizing such oligonucleotides such as routes comprising the synthesis of an intermediate product which may be activated to react with appropriate functional groups such as those of the above-mentioned, preferred embodiments. These methods employ the use of solid supports upon which activation takes place. Such use of the solid support may either be via a DNA synthesizer, by manual manipulation of the solid support or otherwise.

OBJECTS OF THE INVENTION

It is a principal object of the invention to provide oligonucleotide analogs for use in antisense oligonucleotide diagnostics, research reagents, and therapeutics.

It is a further object of the invention to provide nuclease resistant oligonucleotide analogs which possess enhanced cellular uptake.

Another object of the invention is to provide such oligonucleotide analogs which are therapeutically safer and which have greater efficacy than naturally-occurring antisense oligonucleotides.

It is yet another object of the invention to provide methods for synthesis of the modified oligonucleotide using solid supports.

These and other objects will become apparent to persons of ordinary skill in the art from a review of the present specification.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a flow chart of certain synthetic schemes useful for the synthesis of some polyamine functional groups useful in the practice of this invention.

Figure 2 sets forth one method for the synthesis of polyamine conjugated phosphorothioate oligonucleotides useful in this invention.

DETAILED DESCRIPTION OF THE INVENTION

The biological activity of the antisense oligonucleotides previously available has not generally been sufficient for practical therapeutic, research or diagnostic use. This invention directs itself to modified, naturally-occurring oligonucleotides and analogs and methods for effecting such modifications. These modified oligonucleotides and oligonucleotide analogs, exhibit increased biological activity relative to their naturally-occurring counterparts. Furthermore, these modifications may be effected using solid supports which may be manually manipulated or used in conjunction with a DNA synthesizer using methodology commonly known to those skilled in the art.

In the context of this invention, the term "oligonucleotide" refers to a plurality of joined nucleotide units formed in a specific sequence from naturally occurring heterocyclic bases and furanosyl sugar groups joined through the sugar group by native phosphodiester bonds. These nucleotide units may be nucleic acid bases such as guanine, adenine, cytosine, thymine or uracil. The sugar group may be a deoxyribose or ribose sugar. This term refers to both naturally occurring or synthetic species formed from naturally occurring subunits.

"Oligonucleotide analog" as the term is used in connection with this invention, refers to moieties which function similarly to oligonucleotides but which have non naturally occurring portions. Oligonucleotide analogs may

have altered sugar moieties or inter-sugar linkages, for example, phosphorothioates and other sulfur containing species which are known for use in the art. Oligonucleotide analogs may also comprise altered base units or other modifications consistent with the spirit of this invention, in order to facilitate antisense therapeutic, diagnostic or research reagent use of a particular oligonucleotide.

In accordance with the invention, an oligonucleotide sequence is generally chosen which is complementary to DNA or RNA that is involved in the production of proteins whose synthesis is ultimately to be modulated or inhibited in entirety. One preferred embodiment of this invention is an antisense oligonucleotide analog complementary to the DNA or RNA which codes for the *tat* region of HIV. Other preferred embodiments are directed to herpes and papilloma viruses and still others will be apparent to persons of ordinary skill in the art.

Also in accordance with preferred embodiments of this invention, phosphorothioate bonds are substituted for the phosphodiester bonds which normally comprise the sugar phosphate backbone of oligonucleotides. These oligonucleotide analogs are preferably further modified at their terminal 5' end by the addition of a nucleoside analog. This nucleoside analog is most preferably a modified thymidine. Said modified nucleoside can further include the addition of a protecting group such as at the 5' position of the sugar moiety of thymidine. The protecting group in accordance with one preferred embodiment of this invention is a 1,3 diphenylimidazolynyl group.

The modified nucleoside analog, having been incorporated into the oligonucleotide analog, is preferably further modified by the replacement of the protecting group at the 5' position of its sugar moiety with a nitrogenous functional group. The nitrogenous functional group is

preferably a polyamine bearing a terminal functional group such as a primary amine, hydrazine, semicarbazides, thiosemicarbazides or other group capable of reacting with an aldehydic function but which preferably is a polyamine having the structures of polyamine 8 or polyamine 10 as set forth in Figure 1. After reaction of the nitrogenous functional groups with an aldehydic group of the modified nucleoside, the intermediate Schiff bases (imines), hydrazones or semicarbazones are reduced to the substituted amines, hydrazines or semicarbazides. Most preferable the nitrogenous functional group has the structure of polyamine 10 as set forth in Figure 1.

Accordingly, preferred polyamines have the general formula:

$$H_2N(CH_2)_nNH(CH_2)_nNH(CH_2)_nX-$$

wherein X is a primary amine, a hydrazine, a semicarbazide or a thiosemicarbazide and n is an integer between 2 and 6.

Even more preferred polyamines have the general formula:

$$H_2N(CH_2)_nNH(CH_2)_nNH(CH_2)_nNH-$$

wherein n is an integer between 2 and 6. It is preferred that n be 4.

While we do not wish to be bound by theory, the polyamines, which are positively charged, are believed to lie along the negatively charged sugar-phosphate backbone of the oligonucleotide. This configuration is believed to neutralize the negative charges of the oligonucleotide and enhance the cellular penetration of the oligonucleotide. Several recent reports have suggested that a polyamine may form hydrogen bonds to the edges of the base pairs in the major groove, Loke, S.L., Stein, C.A., Zhang, X.H., Mori, K., Nakanishi, M., Subashinghe, G., Cohen, J.S. and Neckers, L.M. *Proc. Natl. Acad. Sci. U.S.A.* 86:3473-3479 (1989). To avoid this problem, this invention preferably does not employ linking the polyamine through a phosphodiester conjugation linkage but instead attaches the polyamine directly to the 5' position. By eliminating the

phosphodiester bond as the conjugation linker and attaching the polyamine directly to the 5' position, it is believed there will be a strong tendency to direct the polyamine chain back along the sugar phosphate backbone of the oligonucleotide conjugate as well as along the backbone of the polyamine-oligonucleotide heteroduplex.

Preferred embodiments of this invention employ a polyamine containing about four carbon units between each amine. Carbon spacers may be arranged in a variety of configurations in relation to the amine groups. This arrangement may be useful in directing the site residence of the polyamine on the oligonucleotide and its heteroduplex. This ability to direct a polyamine to reside at certain positions of the attached oligonucleotides and their heteroduplexes will likely have an important bearing on oligonucleotide uptake and other oligonucleotide properties. Other numbers of carbon unit may be interspaced among the nitrogen atoms of the preferred polyamines in accordance with this invention. Persons of ordinary skill in the art will have wide latitude in selecting optimum configurations for particular circumstances - thus two, five, six or other numbers of carbon units may be employed.

In accordance with certain aspects of the invention, Figure 2 sets forth one chemical synthesis of modified phosphoramidite oligonucleotide analogs. In particular, Figure 2 sets forth novel processes involving the synthesis of 1,3 diphenylimidazolidine protected 5' aldehydic-3'-(2-cyanoethyl-N,N-diisopropylphosphoramidityl) thymidine. This base-stable thymidine analog is placed at the 5' end of a desired oligonucleotide sequence and is activated as needed to couple polyamines such as primary amines, hydrazines, semicarbazides, or thiosemicarbazides.

The starting materials employed by the method set forth in Figure 2 are deoxyribofuranosyl or ribofuranosylnucleosides that can be prepared in high yields by the procedure of, or obvious modification of the

procedure of, Pfitzner and Moffatt, Pfitzner, K.E. et al., J. American Chem. Soc. *85*: 3027 (1963). This procedure describes the oxidation of 3'-acetyl thymidine. The acetyl derivative of the nucleoside to be modified may also be a suitable starting material. Such acetyl derivative may be commercially available or may be prepared by a three step process using procedures known to those skilled in the art.

The first step of this acetylation process is the reaction of t-butyldimethylsilyl chloride (TBDMSCl) with the selected nucleoside to block the nucleoside. Next acetic anhydride (Ac_2O) is added, followed by the addition of Bu_4F to produce the final acetyl derivative of a selected nucleoside. In accordance with the preferred embodiment of this invention, thymidine (the starting material) is converted to 3'-acetyl thymidine using this three step process. Alternatively, commercial 3'-acetyl thymidine is utilized.

The 3'-acetyl nucleoside (**14**) is subsequently oxidized by treatment with DMSO/DCC followed by treatment with 1,2 dianilinoethane to produce 3'-O-acetyl-5'-deoxy-5'-(1,3-diphenylimidazolin-2-yl) nucleoside (**16**). This protecting group has been found to be stable to basic conditions and can be hydrolyzed back to an aldehyde with mild acid. Any of the existing or yet to be discovered groups useful in accomplishing this function may be employed in accordance with the practice of the present invention. In accordance with preferred embodiments 3'-O-acetyl-5'-deoxy-5'-(1,3-diphenylimidazolin-2-yl)thymidine is obtained from the treatment of the 3'-acetyl thymidine with DMSO/DCC followed by treatment with 1,2 dianilinoethane.

Following deblocking of the acetyl group, the protected aldehydic nucleoside is converted to a phosphoramidite (**18**) by standard procedures. Thus the acetyl group is removed by the addition of NH_3/MeOH followed by subsequent phosphitylation by the addition of a phosphityl Cl. The preferred method being phosphitylation

by the addition of 2-cyanoethyl-N,N diisopropylchlorophosphoramidite. Thus in accordance with preferred embodiments 3'-O-acetyl-5'-deoxy-5'-(1,3-diphenylimidazolin-2-yl)thymidine is deblocked followed by conversion to 5'-deoxy-5'-(diphenylimidazolin-2-yl)thymidine 2-cyanoethyl-N,N-diisopropylphosphoramidite.

Separately, a phosphorothioate oligonucleotide having a preselected sequence is extended on a solid support in a 5' direction until such point at which a nucleoside corresponding to the specific modified nucleoside as prepared above need be incorporated into the oligonucleotide sequence. The modified nucleoside is substituted for its naturally occurring terminal nucleoside yielding oligonucleotide analog (20). Most preferably a modified thymidine replaces its naturally occurring counterpart at the 5' end.

Oligonucleotide synthesis may be carried out conveniently through solid state synthesis employing known phosphoramidite methodology on a DNA synthesizer or otherwise. Nucleic acid synthesizers are commercially available and their use is generally understood by persons of ordinary skill in the art as being effective in generating nearly any oligonucleotide of reasonable length which may be desired. Such oligonucleotide chain may be from about 5 to about 50 nucleic acid bases in length. It is more preferred that such functionalities have from 8 to about 40 base units and even more preferred that from about 12 to 20 base units be employed. At present, it is believed that oligonucleotide analogs having about 15 base units will likely be found to be best for the practice of certain embodiments of the present invention.

According to a preferred embodiment of this invention, the *tat* region of HIV is extended in the 5' direction to produce the sequence CCGCCCCTCGCCTCTTGCCT because this region is believed to have the most potent activity in inhibiting the expression of the *tat* protein. See application serial number 521,907, herein incorporated

by reference, filed May 11, 1990 and assigned to the Assignee of this invention. The sequence is then preferably extended three additional nucleotides to TCGCCGCCCCTCGCCTCTTGCCT in order to allow for a T nucleotide on the 5' end. The thymine residue was substituted with a diphenylimidazolinylthymidine as described above.

The resulting phosphoramidite linkage may be preferably sulfurized using the known Beaucage reagent, Beaucage, S.L. et al., *J. Am. Chem. Soc.* 112:1253-1254 (1990), to afford preferred phosphorothioates, or by iodine to afford the phosphodiester linkage. Thus in Figure 2, X is oxygen or sulfur.

The addition of a nitrogenous functional group, preferably a polyamine such as those incorporating terminal primary amines, hydrazines, semicarbazides, or thiosemicarbazides, may be preferably effected once the modified nucleoside has been added to the phosphoramidite oligonucleotide. The first step in this addition is the deprotection of aldehydic groups of the modified nucleoside that has been added to the 5' end of the oligonucleotide. This deprotection can occur, according to preferred embodiments by treatment of the modified nucleoside containing oligonucleotide on the control pore glass (CPG) support (that binds the phosphoramidite oligonucleotide in the synthesizer) with 3% dichloroacetic acid (DCA) in THF. Thereafter, the modified nucleoside containing oligonucleotide on the CPG support is treated with a polyamine to form a Schiff's base (an imine). The Schiff's base is reduced to an amine by the addition of sodium cyanoborohydride ($NaCHBH_3$). Finally the CPG support is treated with ammonium hydroxide to deprotect and remove the polyamine-oligonucleotide conjugate (22) from the support.

Treatment of the CPG support may be effected by a DNA synthesizer or manually using syringes. The preferred functional groups for the embodiment of this invention are polyamines. Most preferably, the polyamine functional

group useful in combination with other preferred components of this invention, is tris(aminobutyl)amine. This polyamine was chosen because of preferred length. The length of the amine-carbon linkages of this group approximately spans the length of a 15 base pair oligonucleotide based upon molecular modeling.

Carbon spacers placed between each amine may be useful in directing the polyamine to reside at certain positions of the attached oligonucleotide. Preferably, carbon spacers will be arranged such that the carbon spacing group will be symmetrical. Four carbon spacers placed between each amine is a preferred embodiment of this invention.

Synthesis of the polyamine group may be performed by procedures known in the art. For preferred embodiments, the synthesis of the polyamine group can be performed according to the steps set forth in Figure 1. Tris(aminobutyl)amine (7) or other like amine can be synthesized by procedures described by Niitsu and Samejima, *Chem. Pharm. Bull.* 34(3):1032-1038 (1986), incorporated by reference herein. Such amines will be protected such as with a t-butyldiphenylsilyl group at one of their primary amino groups according to the procedure set forth by Miller and Braiterman, et al., *Biochemistry* 16:1988-1996 (1977), which involves the addition of TNDPS-Cl and Et_3N. The resulting t-butyldiphenylsilylamine (8) is a convenient amine for reaction with the modified oligonucleotide. More preferably, however, prior to coupling with the modified oligonucleotide compound (8) will be further modified by trifluoroacetylation with trifluoroethyl acetate in Et_2N to produce polyamine (9) followed by selective removal of the t-butyldiphenylsilyl moiety with pyridinium hydrogen fluoride to provide the TFA protected amine (10). This deprotection process is described by Overman, et al, *Tetrahedron Lett.* 27:4391-4394 (1986). The overall process, is amendable to the synthesis of a wide variety of

polyamine in high overall yield. All mentioned references are incorporated by reference herein.

In the alternative, the 5' aldehyde of a nucleoside may be condensed with a trifluoroacetyl protected polyamine. Subsequent reduction of the Schiff's base with sodium cyanoborohydride and phosphitylation of the 3' position will afford a protected polyamine monomer. This protected polyamine monomer may be attached to the 5' end of the oligonucleotide via the DNA synthesizer.

Deprotection and removal of the polyamine-oligonucleotide conjugate from the column may be performed by treatment of the column with ammonium hydroxide. Finally, the composition may be purified using HPLC and gel electrophoresis systems. Such purification procedures are well known by those skilled in the art.

The following examples are illustrative, but not limiting, of the invention.

EXAMPLES

Example 1

1. Preparation of (3'-O-acetyl-1,3 Diphenylimidazolyn-2-yl)nucleoside

3'-O-Acetyl thymidine prepared via standard procedures (1.0 g) was added to a stirred mixture of DCC (3.63 g) and a 1.8 ml aliquot of 0.49 g H_3PO_4 in 5 ml DMSO in DMSO (20 ml) under a nitrogen atmosphere. The mixture was stirred overnight at room temperature. The solvent was removed and the residue partitioned between petroleum ether and water. The organic phase was stripped and the residue dissolved in MeOH (20 ml). Dianilinoethane (0.0047 mM, 1.0 g) was added and the reaction stirred over night to yield 3'-O-acetyl-5'-deoxy-5'-(1,3-diphenylimidazolin-2-yl)thymidine.

2. 3-Phosphoramidite Nucleoside

3'-O-Acetyl-5'-deoxy-5'-(1,3-diphenylimidazolin-2-yl)thymidine is deacetylated with methanolic ammonia and phosphitylated with 2-cyanoethyl-N,N-diisopropylchlorophosphoramidite utilizing using standard

procedures of base deprotection and followed by phosphoramidite phosphitylation to produce 5'-deoxy-5'-(1,3-diphenylimidazolin-2-yl)thymidine 2-cyanoethyl-N,N-diisopropylchlorophosphoramidite.

3. Preparation of Oligonucleotide Sequence

The tat region of an active HIV phosphorothioate oligonucleotide of the sequence 5' CCGCCCCTCGCCTCTTGCCT 3' was built up in the 5' direction utilizing standard automated synthesis on an ABI model 380B DNA synthesizer. Three additional nucleotides were added to extend the sequence to a 5'-thymidine terminal sequence, i.e. the sequence 5' TCGCCGCCCCTCGCCTCTTGCCT 3', with the terminal thymidine nucleotide replaced by 5'-deoxy-5'-(diphenylimidazolin-2-yl)thymidine.

4. Preparation of Polyamine

Tris(aminobutyl)amine is prepared by standard procedures known to those skilled in the art.

5. Addition of Polyamine

The phosphoramidite linked oligonucleotide of the procedure of Example 1-1 will be sulfurized with Beaucage reagent, Beaucage, S.L. et al., *J. Am. Chem. Soc.* 112: 1253-1254 (1990). Next, the CPG support is then treated with 3% dichloroacetic acid in THF to remove the aldehydic blocking group. The CPG support is then subsequently treated with tris(trifluoroacetyl)tris(aminobutyl)amine to form a Schiff's base between the terminal aldehyde and the amine. Sodium cyanoborohydride ($NaCNBH_3$) is then added to reduce the conjugate to an amine. The column is treated with ammonium hydroxide to deprotect and remove the conjugate from column support. The polyamine conjugate is purified with ion exchange HPLC, using a Beckman Gold HPLC system, and with gel electrophoresis.

Example 2

Using the protocol set forth in Example 1, the synthesis of an antisense phosphoramidite oligonucleotide of papilloma virus mRNA cap region, 5'TATGCAAGTACAAAT 3', is performed.

Example 3

Using the protocol set forth in Example 1, the synthesis of an antisense phosphoramidite oligonucleotide of papilloma virus initiation of translation sequence, 5' TCTCCATCCTCTTCACT 3', is performed.

Example 4

Using the protocol set forth in Example 1, the synthesis of an antisense phosphoramidite oligonucleotide sequence of herpes virus, 5' TCATCCATCCTTCGGCC 3', is performed.

Example 5

Using the protocol set forth in Example 1, the synthesis of an antisense phosphoramidite oligonucleotide sequence of herpes virus 5' TGGCCATTTCAACAGA 3', is performed.

Example 6

Using the protocol set forth in Example 1, the synthesis of an antisense phosphoramidite oligonucleotide sequence of herpes virus 5' TCATCCATCCGTCCGCC 3', is performed.

Example 7

Using the protocol set forth in Example 1, the synthesis of an antisense phosphoramidite oligonucleotide sequence of herpes virus 5' TTGGCCATTGGAACCAA 3', is performed.

What is claimed is:

1. An oligonucleotide analog having at least one nucleoside modified by direct attachment to a polyamine.

2. The oligonucleotide analog of claim 1 wherein the oligonucleotide analog ranges in length from about 5 to about 50 nucleotide base units.

3. The oligonucleotide analog of claim 1 wherein said oligonucleotide analog is useful as an antisense therapeutic agent.

4. The oligonucleotide analog of claim 1 wherein said nucleoside is attached at the 5' position of its sugar moiety to the polyamine.

5. The oligonucleotide analog of claim 1 wherein said nucleoside is thymidine.

6. The oligonucleotide analog of claim 1 wherein said polyamine includes an amine, a hydrazine, a semicarbazide, or a thiosemicarbazide functional group thereon.

7. The oligonucleotide analog of claim 1comprising a selected sequence specifically hybridizable with the tat region of a HIV genome.

8. The oligonucleotide analog of claim 7 wherein said selected sequence has a thymidine at its 5' terminal end.

9. The oligonucleotide analog of claim 1 comprising a selected sequence specifically hybridizable with a selected portion of a herpes genome.

10. The oligonucleotide analog of claim 1 comprising a selected sequence specifically hybridizable with a selected portion of a papilloma genome.

11. The oligonucleotide analog of claim 1 wherein said polyamine has the structure:

$H_2N(CH_2)_nNH(CH_2)_nNH(CH_2)_nNH-$

wherein n is an integer between 2 and 6.

12. The oligonucleotide analog of claim 11 wherein n is 4.

13. The oligonucleotide analog of claim 1 comprising at least one phosphorothioate group.

14. A method for synthesizing an antisense oligonucleotide analog wherein at least one nucleoside is modified by direct attachment to a polyamine comprising:

extending an oligonucleotide having a selected sequence of from about 5 to about 50 nucleotide bases in a 5' direction on a solid support to a point in said sequence wherein a selected modified nucleoside is to be incorporated;

incorporating a nucleoside into said oligonucleotide sequence; and

directly attaching a polyamine to the 5' position of said nucleoside.

15. The method of claim 14 wherein:

said polyamine is attached to said nucleoside after said nucleoside is incorporated into said oligonucleotide.

16. The method of claim 14 wherein:

said polyamine is attached to said nucleoside prior to said incorporation of said nucleoside into said oligonucleotide.

17. The method of claim 14 wherein said nucleoside is thymidine.

18. The method of claim 14 wherein said polyamine includes a primary amine, a hydrazine, a semicarbazide, or a thiosemicarbazide functional group thereon.

19. The method of claim 14 wherein said preparing step further comprises the steps of adding a protective group on said nucleoside prior to incorporating said nucleoside in said oligonucleotide; and

deprotecting said nucleoside prior to attachment of said polyamine to said nucleoside.

20. The method of claim 14 wherein said polyamine has the structure:

$H_2N(CH_2)_nNH(CH_2)_nNH(CH_2)_nNH-$

wherein n is an integer between 2 and 6.

21. The method of claim 14 wherein said oligonucleotide is a phosphorothioate oligonucleotide.

22. A method for modulating the production of a protein by an organism comprising contacting the organism with an oligonucleotide analog specifically hybridizable with a selected sequence coding for said protein, wherein at least one nucleoside is modified by direct attachment to a polyamine.

23. The method of claim 22 wherein the oligonucleotide comprises phosphorothioate linkages.

24. The method of claim 22 wherein the oligonucleotide comprises from about 5 to about 50 nucleotide bases.

25. The method of claim 22 wherein said nucleoside is attached at the 5' position of its sugar moiety to the polyamine.

26. The method of claim 22 wherein said nucleoside is thymidine.

27. The method of claim 22 wherein said polyamine includes a primary amine, a hydrazine, a semicarbazide, or a thiosemicarbazide functional group thereon.

28. The method of claim 22 wherein said polyamine has the structure:

$H_2N(CH_2)_nNH(CH_2)_nNH(CH_2)_nNH-$

wherein n is an integer between 2 and 6.

29. The method of claim 28 wherein n is 4.

30. A method for treating an organism having a disease comprising contacting the organism with an oligonucleotide analog specifically hybridizable with a selected sequence of RNA or DNA, wherein at least one nucleoside of the oligonucleotide analog is modified by direct attachment to a polyamine, either alone or in combination with a pharmaceutically acceptable carrier.

31. The method of claim 30 wherein said nucleoside is attached at the 5' position of its sugar moiety to the polyamine.

32. The method of claim 30 wherein said nucleoside is thymidine.

33. The method of claim 30 wherein said polyamine includes a primary amine, a hydrazine, a semicarbazide, or a thiosemicarbazide functional group thereon.

34. The method of claim 30 wherein said selected sequence codes for the tat region of a HIV genome.

35. The method of claim 30 wherein said selected sequence has a thymidine at its 5' terminal end.

36. The method of claim 30 wherein said selected sequence codes for a selected portion of a herpes genome.

37. The method of claim 30 wherein said selected sequence codes for a selected portion of a papilloma genome.

38. The method of claim 30 wherein said polyamine has the structure:

$H_2N(CH_2)_nNH(CH_2)_nNH(CH_2)_nNH-$

wherein n is an integer between 2 and 6.

39. The method of claim 38 wherein n is 4.

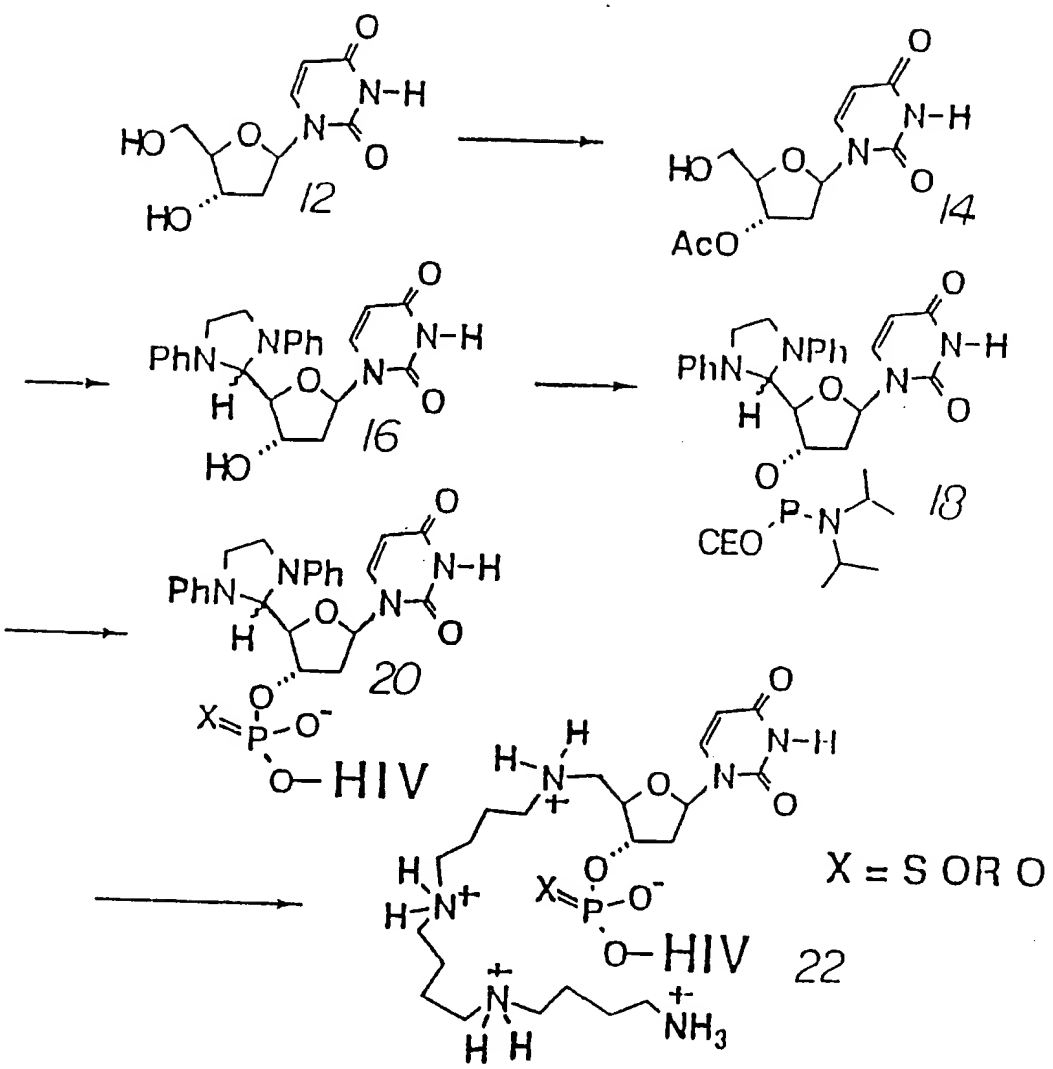

FIGURE 2

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US91/04086

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) [6]

According to International Patent Classification (IPC) or to both National Classification and IPC

IPC(5): C07H 17/00; A61K 31/78; C12P 21/06, 21/04
U.S. CL.: 536/27; 514/44; 435/69.1, 71.1

II. FIELDS SEARCHED

Minimum Documentation Searched [7]

Classification System	Classification Symbols
U.S.	536/27; 514/44; 435/69.1, 71.1

Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched [8]

III. DOCUMENTS CONSIDERED TO BE RELEVANT [9]

Category *	Citation of Document, [11] with indication, where appropriate, of the relevant passages [12]	Relevant to Claim No. [13]
X Y	Nucleic Acid Research, Volume 17, No. 18, issued 1989. Nelson et al., "A new and versatile reagent for incorporation multiple primary aliphatic amines into synthetic oligonucleotides", pages 7179-7186, particularly pages 7183 and 7184, and the abstract.	1-4, 6 & 13 5 & 7-21
Y	Proceedins of the National Academy of Science, Volume 84, issued February 1987, Lemaztre, et al., "Specific antiviral activity of a Poly(L-lysine)-conjugated oligo-dexoyribonucleotide sequence complementary to vesicular stomatitis virus N protein mRNA initiation site" pages 648-652, see entire document.	14-39

* Special categories of cited documents: [10]

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search	Date of Mailing of this International Search Report
06 November 1991	11 DEC 1991
International Searching Authority	**Signature of Authorized Officer**
ISA/US	John W. Rollins ebw

Form PCT/ISA/210 (second sheet) (Rev.11-87)

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)

Category *	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No
Y	Biochemistry, Volume 27, issued 1988, Wu et al., "Evidence for target gene delivery to Hep G_2 Hepatoma Cells In vitro", pages 887-892, see entire documents.	14-39